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ARTICLE

Risk Assessment for Salmonellosis in Chicken in South Korea: The Effect of Salmonella Concentration in Chicken at Retail

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Tel: +82-2-450-4121 Fax: +82-2-3436-4128 E-mail: bracstu3@konkuk.ac.kr Abstract Salmonellosis caused by chicken consumption has been a critical issue in food safety worldwide, including in Korea. The probability of illness from consumption of chicken was simulated in study, based on the recipe of Dakgalbi, a commonly eaten chicken dish in Korea. Additionally, the processing stage at slaughterhouses to decrease Salmonella concentration in broilers was modeled to explore its effect on the likelihood of illness. A Monte Carlo simulation model was created using @RISK. Prevalence of Salmonella in chickens at the retail stage was found to be predominantly important in determining the probability of illness. Other than the prevalence, cooking temperature was found to have the largest impact on the probability of illness. The results also demonstrated that, although chlorination is a powerful tool for decreasing the Salmonella concentration in chicken, this effect did not last long and was negated by the following stages. This study analyzes the effects of variables of the retail-to-table pathway on the likelihood of salmonellosis in broiler consumption, and also evaluates the processing step used to decrease the contamination level of Salmonella in broilers at slaughterhouses. According to the results, we suggest that methods to decrease the contamination level of Salmonella such as chlorination had little effect on decreasing the probability of illness. Overall, these results suggest that preventing contamination of broiler with Salmonella must be a top priority and that methods to reduce the concentration of Salmonella in broilers at slaughterhouses hardly contribute to safe consumption of Salmonella-contaminated chicken.

Keywords predictive microbiology, risk analysis, chicken, Salmonella, quantitative microbial risk assessment

Introduction

Establishment of food safety is a major task for securing the well-being of a population and remains a public health challenge. Microorganisms are the major cause of food poisoning incidents (Nyachuba, 2010), and the long-standing and continuous outbreaks of food poisoning by microorganisms demonstrate the difficulty of controlling pathogenic microorganisms. For example, 4,265 cases of food poisoning were reported in Korea between 2002 and 2017, affecting 111,947 patients (Portal of Food Safety Information, 2018). Among them, 332 cases involving 10,378 patients were attributed to *Salmonella* infection, which was the third most common cause of food poisoning only after enteropathogenic *Escherichia coli* and norovirus (Portal of Food Safety Information, 2018). Survey studies have shown that poultry consumption is one of the major causes of *Salmonella* infection in Korea (Chung et al., 2003; Kim et al., 2012).

Oscar (2004) performed a risk analysis for *Salmonella* on broilers along the pathway from retail to consumption. However, it is necessary to explore the link between the processing at slaughterhouses and the pathway from retail to consumption because the likelihood of illness after consumption depends on the concentration of *Salmonella* in broilers at retail (Oscar, 2004). Additionally, evaluation of the impact of the stage of production on risk management emphasized the significance of the processing stage of broilers for food safety (Nauta et al., 2005). It is, thus, necessary to understand how much contamination can be managed in the stages from retail to consumption, and how the level of contamination can be minimized in the stage of processing at slaughterhouses. Chlorination has long been used to decrease the *Salmonella* contamination level in poultry (Paul et al., 2017). A chlorination level of 20 ppm is widely used to maintain food safety in poultry slaughterhouse in Korea (Woo, 2007). However, Woo (2007) found that treating chickens with chlorination did not have a significant effect on reducing the *Salmonella* load through a survey on the status of microbial contamination in slaughterhouses. It is, therefore, necessary to determine whether a significant increase in chlorine concentration can provide the required reduction in the concentration of *Salmonella* in chicken.

Quantitative microbial risk assessment (QMRA) is a relatively useful method that was introduced to quantify risk (WHO, 2002). With the use of QMRA, the probability of acquiring a microbial infection can be estimated by quantifying the prevalence and concentrations of microbes present during each procedure related to food preparation and consumption (Whiting and Buchanan, 1997). Oscar (2004) built QMRA for *Salmonella* and whole chickens to determine the relationship between the most probable number of *Salmonella* cells and the illness probability, and how this relationship is affected by each stage in the retail-to-table pathway in the United States. Following this work, in the present study, we built QMRA tracking the procedures from the retail to household consumption stages with respect to *Salmonella* in chicken. given the specific processing conditions and data of Korea, and explored hypothetical situations in which illness caused by consumption of *Salmonella*-contaminated chicken is more likely.

QMRA of *Salmonella* in broilers should be region-specific from country to country, depending on the contamination level of *Salmonella* in broilers at retail shops and the circumstances of shopping and cooking styles. We built the QMRA based on the Korean circumstance and chose the cooking method of Dakgalbi, which is a traditional and common recipe for chickens in Korea (Muhlisin et al., 2012).

The goal of the study was two-fold: (1) to conduct a baseline microbial risk assessment for *Salmonella* in chicken employing Monte Carlo simulation, and to perform a sensitivity analysis for the parameters on the probability of illness; (2) to explore how the level of chlorination affects the likelihood of illness. Based on the results, we provide a potential foundation for food safety in terms of risk management for *Salmonella* in chicken.

Materials and Methods

QMRA design

We modeled the retail-to-table pathway in Korea by covering the steps beginning at purchase of a chicken from a retail

shop to the final consumption of the chicken. Accordingly, the QMRA consisted of the following four stages: stage 1, initial contamination level at retail; stage 2, microbial growth during transport from retail shops to household refrigerators; stage 3, thermal inactivation during cooking; and stage 4, dose-dependent illness after consumption. In addition, the effects of the chlorination procedure were considered at stage 1, although in practice chlorination is actually performed during the chilling process in slaughterhouses (James et al., 2006). *Salmonella* was not assumed to grow during storage in a refrigerator, because properly working refrigerators keep the temperature below 10°C, i.e., the minimum growth temperature of *Salmonella* (WHO, 2002). The model excluded the potential of cross-contamination of *Salmonella* between chicken and other ingredients such as vegetables so as to best focus on the salmonellosis caused specifically by chicken consumption, due to the lack of survey data for cross-contamination in Korea. We tried to incorporate realistic consumption conditions in Korea into the model as much as possible. If relevant data were not available, we referred to foreign data, expert opinion, and logical inferences. The model was simulated with @RISK (Palisade Corporation, Ithaca, NY) using a Monte Carlo simulation with settings of Latin hypercube sampling and was iterated 100,000 times. We employed a high number of iterations because of the extremely low values of the likelihood of illness estimated by this study.

Stage 1: Retail

The first stage in the QMRA simulated the contamination level of *Salmonella* in chicken at a retail shop using the prevalence and concentration of *Salmonella* in chicken. The prevalence of *Salmonella* in retail chicken in Korea is widely variable, with values of 22.4% (Kim et al., 2012), 36.0% (Chung et al., 2003), and 42.3% (Hyeon et al., 2011) reported. We used the minimum (22.4%), maximum (42.3%), and average of the four values (31.65%). The concentration of *Salmonella* per chicken was estimated based on a previous survey conducted on whole raw chicken purchased from a local market in Korea (Hong et al., 2008).

Stage 2: Transport

The second stage simulated the growth of Salmonella in chicken during transportation of the chicken from the retail shop to the home. The period of transport was estimated by using the locations of "e-mart", a major retail franchise in Korea. We spotted two locations of e-mart in Seoul, Korea, and measured the distance between the two, which was 4.1 km. We supposed that consumers visit an e-mart at half of this distance (2.1 km) using a private car or public transportation. A private car and public transportation were expected to take 17 and 35 min, respectively. We used the minimum (17 min), maximum (35 min), and average (26 min) for the Pert distribution. The surveyed average shopping time of Koreans was 1–2 hours (Trendmonitor, 2018). We added half of the average shopping time to the transport time. The ambient temperature during transport was assumed to be the average temperature in Korea (Korea Meteorological Administration, 2018). The transport temperature was used to calculate the growth rate and lag time to estimate the contamination level during transport. The temperature of raw chicken purchased at a retail shop was supposed to increase exponentially per 60/min. Until the temperature of the chicken reached ambient temperature, the average of temperature of the chicken at retail and the temperature of the chicken after transport were assumed to be the temperature of the chicken. If the temperature of the chicken reached the ambient temperature during transport, the temperature of the chicken was assumed to be the ambient temperature. The growth rate of Salmonella in chicken was determined based on the secondary model of Baranyi (Juneja et al., 2007), and the lag time was based on experimental validation of the lag time of Salmonella Typhimurium (Oscar, 2005). The maximum theoretical concentration of 9.5 Log CFU/g was assumed based on the findings of Silva et al. (2009).

Stage 3: Cooking

The third stage simulated the thermal inactivation of *Salmonella* during the cooking of chicken. The cooking method was based the recipe of Dakgalbi. The parameters included in the cooking stage were cooking temperature and time. The chicken was cooked at 75°C to reach an internal temperature of 71°C (Song et al., 2014), such that 0.8 and 1.2 times 71°C were used for the Pert distribution. We referred to Lee et al. (2011) for cooking time. The cooking temperature was used to calculate the decimal reduction time (D-value), which is the time required to kill 90% of the microbes of interest (Liu et al., 2015). A thermal inactivation model developed for *Salmonella* in chicken patties was used to estimate changes in the D-value depending on temperature (Murphy et al., 2002). The ratio of cooking time to the D-value was used to calculate the log cycle reduction, which is the overall parameter indicating the thermal inactivation effect at the cooking stage.

Stage 4: Consumption

The fourth and final stage simulated the consumption of chicken. The amount of chicken consumed was set to reflect the typical amount of Dakgalbi eaten by Koreans in a given meal. The dose-response model of *Salmonella* referred to WHO (2002). The outcome of this QMRA was the likelihood of salmonellosis caused by consumption of *Salmonella*-contaminated chicken.

Baseline model

In the baseline model, we used the most probable ranges of parameter values that may affect the risk of salmonellosis. The effect of chlorination to decrease the concentration of *Salmonella* was not considered in the baseline model. The formulas, units, and input settings used in the baseline model are shown in Table 1. In addition, sensitivity analysis was performed to provide a quantitative measure of the most important parameters affecting the probability of illness from *Salmonella* via the pathway explored in the baseline model. The value of performing this sensitivity analysis is that the general results can be used to prioritize risk mitigation strategies based on the parameters that emerge to show the strongest impact on risk (Zwietering and van Gerwen, 2000). Spearman rank correlation using @RISK was employed to determine the parameters that are most highly correlated with the probability of illness from *Salmonella*.

Table 1. Parameters used in the baseline model

Unit operation	Variables	Description	Unit	Formula	Reference
Retail	P	Prevalence of Salmonella at retail		Pert(22.4, 31.65, 42.3)	(Chung et al., 2003; Hyeon et al., 2011; Kim et al., 2012)
	CO_{SC}	Contamination of chicken with Salmonella at retail		Discrete([0, 1], [100–P, P])	
	Te _R	Temperature of chicken at retail	$^{\circ}\!\mathrm{C}$	4	(WHO, 2002)
	LC_{SC}	Log concentration of Salmonella at retail	Log(CFU/g)	Pert(4.3, 5.3, 6.3)	(Hong et al., 2008)
	C_{SC}	Concentration of Salmonella at retail	CFU/g	$10^{\wedge}LC_{SC}$	
	C_{CH}	Concentration of chlorination	ppm	0	
	λ	Parameter of Salmonella reduction effect		1	
	O_{RE}	Contamination level at retail	CFU/g	IF($CO_{SC}=0, 0, C_{SC}\times\lambda$)	

Table 1. Parameters used in the baseline model (continued)

Unit operation	Variables	Description	Unit	Formula	Reference
Transport	Ti_{T}	Transport and shopping time	h	Pert(17/60, 26/60, 35/60)+Pert(0.5, 0.75, 1)	(Trendmonitor, 2018)
	Тет	Ambient temperature	$^{\circ}\!\mathrm{C}$	Pert(min, median, max)	(Korea Meteorological Administration, 2018
	Te _{TR}	Chicken temperature after transport	°C	$IF(exp(Ti_T)-1+Te_R>Te_T, Te_T, exp(Ti_T)-1+T_{eR})$	
	AvTe _{TR}	Average temperature between retail and after transport	$^{\circ}\mathrm{C}$	Average(TeR, TeTR)	
	$TIAM_{TR}$	Time when chicken temperature reaches ambient temperature	h	$LOG(Te_T, exp(1))$	
	LT	Lag time of Salmonella	h	$1/(0.02082 \times \text{AvTe}_{\text{TR}} - 0.01235)^2$	(Oscar, 2005)
	GR _{RETR}	Growth rate of Salmonella between retail and transport before chicken reaches ambient temperature	Log(CFU/g/h)	$IF(AvTe_{TR}>TeT, \\ 0.0019\times(AvTe_{TR}- \\ 3.35)^2\times(1- \\ exp(0.29\times(AvTe_{TR}- \\ 48.01))), 0)$	(Juneja et al., 2007)
	GR _{AMTR}	Growth rate of <i>Salmonella</i> at ambient temperature during transport	Log(CFU/g/h)	0.0019×(Te _T -3.35) ² ×(1- exp(0.29×(Te _T -48.01)))	(Juneja et al., 2007)
	MPD	Maximum population density	Log(CFU/g)	9.5	(Silva et al., 2009)
	GS	Growth of Salmonella	Log(CFU/g)	$\begin{split} & IF(TI_T \!\!<\! TIAM_{TR}, \\ & GR_{RETR} \!\!\times\! Ti_T, \\ & GR_{RETR} \!\!\times\! TIAM_{TR} \!\!+\! GR_{AMTR} \\ & \times\! (Ti_T \!\!-\! TIAM_{TR})) \end{split}$	
	O_{TR}	Contamination level after transport	CFU/g	IF(O _{RE} +exp(GS) <exp(mp D), O_{RE}+exp(GS), exp(MPD))</exp(mp 	
Cooking	Te _C	Cooking temperature	°C	Pert(71×0.8, 71, 71×1.2)	(Song et al., 2014)
	Ti_{C}	Cooking time	min	Pert(10, 15, 20)	(Lee et al., 2011)
	D	D-value	min	10^(8.7344-(0.1316×Te _C))	(Murphy et al., 2002)
	LCR	Log cycle reduction	-	Ti _C /D	
	Oc	Contamination level after cooking	CFU/g	$(10^{-1}(-LCR)) \times O_{TR}$	
Consumption	CW	Consumption weight	g	Pert(100, 200, 300)	(Fatsecret, 2018)
	Dose	Amount of Salmonella consumed	CFU	$O_C \times CW$	
	P_{ILL}	Probability of illness		IF(P=0, 0, (1- (1+10^Dose/2885)^(- 0.3126)))	(WHO, 2002)

Chlorination model

In the chlorination model, we applied the different levels of chlorination effects in stage 1 to simulate the reduction of *Salmonella* concentration in chickens at retail, and we explored whether the decrease in *Salmonella* concentration in chickens at retail by chlorination can affect the likelihood of illness after consumption of *Salmonella*-contaminated chicken. A predictive

model for the effect of chlorination to reduce the concentrations of *Salmonella* in chicken was based on previous findings showing that 50, 100, and 200 ppm chlorine reduced the *Salmonella* load by 1 Log CFU/g (Northcutt et al., 2005), 1.4 Log CFU/g (Hong et al., 2008), and 2 Log CFU/g (Lu and Wu, 2012), respectively. The square root of the published chlorination effect was plotted against the chlorination concentration and linear regression was used to model the effect of chlorine concentration on *Salmonella* reduction (Fig. 1). In the chlorination model, we first used fixed values of chlorination concentration (0, 50, 100, 200, and 400 ppm). Then, for sensitivity analysis, we used the Pert distribution of chlorination concentration of 0, 200, and 400 ppm for minimum, most likely, and maximum values, respectively.

Results

Baseline model

The simulation results of the baseline models showed that the likelihood of illness was not affected by the transport temperature (Table 2). The short distances between retail shops in Seoul contributed to ensure food safety during transport, because microbial growth could hardly be achieved during such a short time of exposure to ambient temperature.

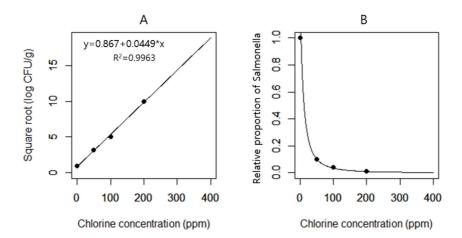


Fig. 1. (A) Reciprocal of the square root of *Salmonella* reduction in chicken versus chlorine concentration. (B) Parameter (λ) of *Salmonella* reduction depending on chlorination concentration. The concentration of *Salmonella* in chicken meat after chlorination was calculated by multiplying λ with the concentration of *Salmonella* in chicken. Dots represent published data.

Table 2. Likelihood of illness in each month

Month	Average likelihood of illness
March	0.0127
April	0.0112
May	0.0122
June	0.0124
July	0.0111
August	0.0120
September	0.0122
October	0.0120
November	0.0125

The sensitivity analysis indicated that the prevalence of *Salmonella* in broilers had a predominant impact on deciding the likelihood of illness after consumption of Dakgalbi (Fig. 2A). We also performed the sensitivity analysis with the deterministic value of *Salmonella* prevalence, which meant that the chicken at retail was always contaminated with *Salmonella* (Fig. 2B). The results of this sensitivity analysis showed that cooking temperature had a predominant effect on deciding the probability of illness.

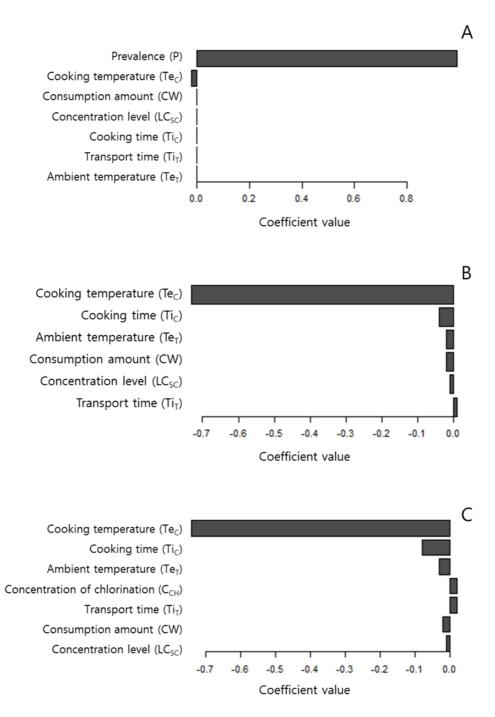


Fig. 2. Tornado chart of sensitivity analysis for the probability of illness. (A) Sensitivity analysis for the results of August in the baseline model. (B) Sensitivity analysis for the results of August in the baseline model with the deterministic value of prevalence. (C) Sensitivity analysis for the results of the chlorination model in August.

Chlorination model

In the chlorination model, we explored whether the effect of chlorination on reduction of *Salmonella* concentration in broiler chickens can reduce the probability of illness. Increased concentration of chlorination had little effect on the probability of illness (Fig. 3). This finding indicates that the usage of methods including chlorination during chilling at poultry slaughterhouses that could decrease the concentration of *Salmonella* should not be recommended for reducing the probability of illness. The usage of chlorination during the chilling process was simulated to reduce the concentration of *Salmonella* in chickens at the retail stage. However, the sensitivity analysis of the chlorination model showed that the effect of the reduced concentration through chlorination was negated by the following stages, particularly the cooking stage (Fig. 2C).

Discussion

The QMRA developed in this study covers all of the processes involved in the likelihood of becoming infected with *Salmonella* from a contaminated chicken, from the retail to consumption stage. By combining published data for the *Salmonella* contamination level (Hong et al., 2008) and prevalence (Chung et al., 2003; Hyeon et al., 2011; Kim et al., 2012) at retail in Korea, reduction of *Salmonella* concentration by chlorination (Hong et al., 2008; Lu and Wu, 2012; Northcutt et al., 2005), microbial growth (Juneja et al., 2007; Oscar, 2005; Silva et al., 2009), thermal inactivation (Murphy et al., 2002), and dose dependence of food illness (WHO, 2002), we estimated the probability of illness caused by the consumption of *Salmonella*-contaminated chicken and explored the relative significance of each parameter for the probability of illness. In this way, we were able to determine the parameters that should be focused on for preventing illness.

The methods to reduce the concentration of *Salmonella* in broilers during processing steps at slaughterhouses could reduce the concentration of *Salmonella* on raw chickens at retail shops, but this effect was found to be negligible compared to the effect of cooking for thermal inactivation of *Salmonella*. Although increasing the concentration of chlorination can decrease the probability of illness to some extent at the retail stage, the probability of illness was found to be minimally affected by the chlorination concentration. This finding emphasizes that disinfection of microbes at slaughterhouses should be performed very meticulously after sufficient validation is found. In addition, a high concentration of chlorination is not recommended because a chlorine-based washing solution might be detrimental to human health or deteriorate the quality of chicken (Lu and Wu, 2012).

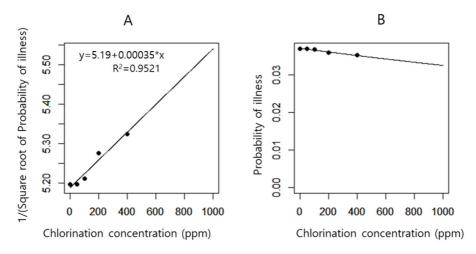


Fig. 3. (A) Reciprocal of the square root of the probability of illness versus chlorine concentration. (B) Probability of illness depending on chlorination concentration. Dots represent the simulation results of the chlorination model.

The finding that the prevalence of Salmonella in chicken has a predominant impact on the likelihood of illness suggests the importance of prevention of Salmonella contamination at stages of rearing and slaughtering. The efforts to decrease the contamination level of Salmonella in chicken at the slaughtering stage should be preceded by efforts to prevent contamination. Therefore, it is necessary to understand the routes of Salmonella infection in poultry rearing farms and to prevent the contamination of broiler with Salmonella (Rose et al., 1999). In slaughterhouses, efforts to decrease the contamination of chicken with microbial pathogens should be focused on the steps that may cause cross-contamination during slaughtering, such as evisceration rather than the chilling step that uses disinfectant agents (Hue et al., 2010). Specifically, to reduce the prevalence of Salmonella in chicken meat at retail, it is essential to determine the points of the process that are most prone to Salmonella contamination. Kim et al. (2012) showed that the highest prevalence of Salmonella occurred in broiler transporting trucks (71.43%), followed by the slaughterhouse (63.89%) and broiler farms (16.05%). According to this finding, further efforts to reduce the prevalence at retail should be concentrated on the processes involved in the transportation and slaughter of broilers. In addition, the prevalence of Salmonella in chicken meat will vary substantially depending on the distribution channel. One survey in Korea demonstrated that the prevalence of Salmonella in chicken meat was 25% at wholesale markets, 44.4% at retail stores, and 55.56% at traditional markets (Hyeon et al., 2011). This finding emphasizes the need to apply strict food safety protocols, particularly at traditional markets. Comparison to studies on the prevalence of Salmonella in chicken meat at retail worldwide shows that the prevalence in Korea is similar to that reported in other countries: 41.6% (Zhu et al., 2014) and 33.8% (Huang et al., 2016) in China, 30% in the USA (Oscar, 2004), and 30% in Canada (Smadi and Sargeant, 2012).

Microbes are sensitive to high temperature (van Asselt and Zwietering, 2006); thus, encouraging the public to cook meat at a high temperature can reduce the possibility of an outbreak of foodborne illness. The sensitivity analysis without Salmonella prevalence (Fig. 2B) supported the well-known fact that even a slight change in cooking temperature can generate a massive impact on the probability of illness, even without changing any other parameter (van Boekel, 2002). Nevertheless, it is not always easy to apply a sufficiently high temperature to chicken meat, because some widely eaten chicken recipes in Korea make it difficult to effectively cook the deeper parts of the chicken meat. Compared to boiling and frying, stir-frying in a pan is a poor condition for achieving high internal temperature of meat pieces and for accomplishing effective thermal inactivation of Salmonella. For example, Koreans often use built-in grills or portable burners on tables to stir-fry chicken meat while eating it. This cooking style increases the likelihood of eating less-cooked chicken meat and the likelihood of causing cross-contamination from uncooked chicken meat to vegetables because the meat is cooked not by a professional chef but by consumers. Therefore, the risk caused by the consumption of less-cooked chicken meat is particularly significant due to this characteristic Korean cooking style. Additionally, one recipe in Korea and Japan involves eating chicken sashimi, and this eating habit has been involved in food safety concerns (Asoshima et al., 2015; Moore and Matsuda, 2007). Considering the critical importance of cooking and the limited effect of chlorination on the probability of illness, eating raw chicken meat should be strongly discouraged in terms of food safety, unless the prevalence and concentration of Salmonella in raw chicken at retail are substantially improved.

In this study, we excluded some parameters that may also affect the probability of illness but cannot be logically assumed or reliably referred. For example, cross-contamination between raw chicken and vegetables during the preparation of foods may occur in the kitchen and can be an important source of salmonellosis (Luber, 2009). The contamination of *Salmonella* from a contaminated raw chicken to a cooked chicken is also possible and is called self-contamination (Oscar, 2004). However, this was too difficult to simulate as there are too many factors related to such cross-contamination, which would be

difficult to model with only a few parameters (for example, the transfer rate from raw chicken to vegetables or utensils, consumption amount of vegetables, microbial growth rate on vegetables and utensils, and storage of contaminated vegetables). Although Oscar (2004) applied data from consumer surveys of handling practices of cooked food and laboratory studies that quantified the transfer rates of *Salmonella*, we did not refer to these data in the present model, because such variables would be highly dependent on the specific recipe and conventional handling practices of chicken in different countries. Therefore, there is a need to properly quantify the level of cross-contamination from chicken in Korea, especially given the increasing trend of eating chicken with uncooked vegetables such as chicken salad.

Conclusion

In summary, given the current data on the concentration of *Salmonella* in chicken at the retail level, one risk management option might be to prioritize the prevention of *Salmonella* contamination. Other than reducing the prevalence, use of a high cooking temperature is the most reliable method to ensure safe consumption of chicken without illness caused by *Salmonella*. Although methods to reduce the concentration of *Salmonella* at slaughterhouses can be an effective measure to decrease the *Salmonella* concentration in chicken at retail, as a sole practice, these methods are far from guaranteeing safety, even at high concentrations of chlorination.

The establishment of standardized chicken risk management guidelines would be useful for consumers concerned about individual or family food safety. In this regard, the present risk assessment may also be useful for food safety-related authorities when setting a quantitative standard for *Salmonella* in chicken. Finally, this risk assessment points to the need for additional research on the effects of other factors (e.g., concentration of *Salmonella* on chicken, cross-contamination, and effects of cooking depending on the recipe), which may be useful in securing the safety of chicken from salmonellosis.

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References

- Asoshima N, Matsuda M, Shigemura K, Honda M, Yoshida H, Oda T, Hiwaki H. 2015. Isolation of *Escherichia albertii* from raw chicken liver in Fukuoka City, Japan. Jpn J Infect Dis 68:248-250.
- Chung YH, Kim SY, Chang YH. 2003. Prevalence and antibiotic susceptibility of *Salmonella* isolated from foods in Korea from 1993 to 2001. J Food Prot 66:1154-1157.
- Fatsecret. 2018. Available from: www.fatsecret.kr/%EC%B9%BC%EB%A1%9C%EB%A6%AC-%EC%98%81%EC%96% 91%EC%86%8C/%EC%9D%BC%EB%B0%98%EB%AA%85/%EB%8B%AD%EA%B0%80%EC%8A%B4%EC%82% B4?portionid=9981348&portionamount=1.000. Accessed at Jan 1, 2018.
- Hong Y, Ku K, Kim M, Won M, Chung K, Song KB. 2008. Survival of *Escherichia coli* O157: H7 and *Salmonella typhimurium* inoculated on chicken by aqueous chlorine dioxide treatment. J Microbiol Biotechnol 18:742-745.
- Huang J, Zong Q, Zhao F, Zhu J, Jiao XA. 2016. Quantitative surveys of *Salmonella* and *Campylobacter* on retail raw chicken in Yangzhou, China. Food Control 59:68-73.

- Hue O, Le Bouquin S, Laisney MJ, Allain V, Lalande F, Petetin I, Rouxel S, Quesne S, Gloaguen PY, Picherot M, Santolini J, Salvat G, Bougeard S, Chemaly M. 2010. Prevalence of and risk factors for *Campylobacter* spp. contamination of broiler chicken carcasses at the slaughterhouse. Food Microbiol 27:992-999.
- Hyeon JY, Chon JW, Hwang IG, Kwak HS, Kim MS, Kim SK, Choi IS, Song CS, Park C, Seo KH. 2011. Prevalence, antibiotic resistance, and molecular characterization of *Salmonella* serovars in retail meat products. J Food Prot 74:161-166.
- James C, Vincent C, de Andrade Lima T, James SJ. 2006. The primary chilling of poultry carcasses—A review. Int J Refrig 29:847-862.
- James WO, Brewer RL, Prucha JC, Williams Jr W, Parham DR. 1992. Effects of chlorination of chill water on the bacteriologic profile of raw chicken carcasses and giblets. J Am Vet Med Assoc 200:60-63.
- Juneja VK, Melendres MV, Huang L, Gumudavelli V, Subbiah J, Thippareddi H. 2007. Modeling the effect of temperature on growth of *Salmonella* in chicken. Food Microbiol 24:328-335.
- Kim MS, Lim TH, Jang JH, Lee DH, Kim BY, Kwon JH, Choi SW, Noh JY, Hong YH, Lee SB, Yang SY, Lee HJ, Lee JB, Park SY, Choi IS, Song CS. 2012. Prevalence and antimicrobial resistance of *Salmonella* species isolated from chicken meats produced by different integrated broiler operations in Korea. Poult Sci 91:2370-2375.
- Korea Meteorological Administration. 2018. Temperature Database of Seoul. Available from: http://www.kma.go.kr/weather/climate/average_30years.jsp?yy_st=2011&stn=108&norm=M&x=12&y=10&obs=0&mm=5&dd=15. Accessed at Jan 1, 2018.
- Lee SK, Choi WH, Muhlisin M, Kang SM, Kim CJ, Ahn BK, Kang CW. 2011. Quality comparison of Chuncheon dakgalbi made from Korean native chickens and broilers. Korean J Food Sci An 31:731-740.
- Liu H, Gong J, Chabot D, Miller SS, Cui SW, Ma J, Zhong F, Wang Q. 2015. Protection of heat-sensitive probiotic bacteria during spray-drying by sodium caseinate stabilized fat particles. Food Hydrocoll 51:459-467.
- Lu Y, Wu C. 2012. Reductions of *Salmonella enterica* on chicken breast by thymol, acetic acid, sodium dodecyl sulfate or hydrogen peroxide combinations as compared to chlorine wash. Int J Food Microbiol 152:31-34.
- Luber P. 2009. Cross-contamination versus undercooking of poultry meat or eggs—Which risks need to be managed first? Int J Food Microbiol 134:21-28.
- Moore JE, Matsuda M. 2007. Consumption of raw chicken sashimi, Kyushu Island, Japan—Risk of campylobacteriosis or not? Travel Med Infect Dis 5:64-65.
- Muhlisin M, Kang SM, Choi WH, Kim CJ, An BK, Kang CW, Lee SK. 2012. New approach to Chuncheon Dakgalbi processing by various chicken materials, seasoning and cooking methods. Korean J Food Sci An 32:740-748.
- Murphy RY, Duncan LK, Johnson ER, Davis MD, Smith JN. 2002. Thermal inactivation *D*-and *z*-values of *Salmonella* serotypes and *Listeria innocua* in chicken patties, chicken tenders, franks, beef patties, and blended beef and turkey patties. J Food Prot 65:53-60.
- Nauta M, van Der Fels-Klerx I, Havelaar A. 2005. A poultry-processing model for quantitative microbiological risk assessment. Risk Anal 25:85-98.
- Northcutt JK, Smith DP, Musgrove MT, Ingram KD, Hinton Jr A. 2005. Microbiological impact of spray washing broiler carcasses using different chlorine concentrations and water temperatures. Poult Sci 84:1648-1652.
- Nyachuba DG. 2010. Foodborne illness: Is it on the rise? Nutr Rev 68:257-269.
- Oscar TE. 2005. Validation of lag time and growth rate models for Salmonella Typhimurium: acceptable prediction zone

- method. J Food Sci 70:129-137.
- Oscar TP. 2004. A quantitative risk assessment model for Salmonella and whole chickens. Int J Food Microbiol 93:231-247.
- Paul NC, Sullivan TS, Shah DH. 2017. Differences in antimicrobial activity of chlorine against twelve most prevalent poultry-associated *Salmonella* serotypes. Food Microbiol 64:202-209.
- Portal of Food Safety Information. Statistics of food poisoning. Available at: https://www.foodsafetykorea.go.kr/main.do. Accessed at Jan 1, 2018.
- Rose N, Beaudeau F, Drouin P, Toux JY, Rose V, Colin P. 1999. Risk factors for *Salmonella enterica* subsp. *enterica* contamination in French broiler-chicken flocks at the end of the rearing period. Prev Vet Med 39:265-277.
- Silva RR, Moraes CA, Bessan J, Vanetti MCD. 2009. Validation of a predictive model describing growth of *Salmonella* in enteral feeds. Braz J Microbiol 40:149-154.
- Smadi H, Sargeant JM. 2012. Quantitative risk assessment of human salmonellosis in Canadian broiler chicken breast from retail to consumption. Risk Anal 33:232-248.
- Song YR, Kim DS, Muhlisin M, Seo TS, Jang A, Pak JI, Lee SK. 2014. Effect of chicken skin and pork backfat on quality of dakgalbi-taste chicken sausage. Korean J Poult Sci 41:181-189.
- Trendmonitor. 2018. Available at: www.trendmonitor.co.kr/tmweb/trend/allTrend/detail.do?bIdx=574&code=0201&trendType =CKOREA. Accessed at Jan 1, 2018.
- van Asselt ED, Zwietering MH. 2006. A systematic approach to determine global thermal inactivation parameters for various food pathogens. Int J Food Microbiol 107:73-82.
- van Boekel MAJS. 2002. On the use of the weibull model to describe thermal inactivation of microbial vegetative cells. Int J Food Microbiol 74:139-159.
- Whiting RC, Buchanan RL. 1997. Development of a quantitative risk assessment model for *Salmonella enteritidis* in pasteurized liquid eggs. Int J Food Microbol 36:111-125.
- WHO [World Health Organization]. 2002. Risk assessments of *Salmonella* in eggs and broiler chickens. Food & Agriculture Org, Geneva, Switzerland.
- Woo YK. 2007. Survey on the status of microbial contamination of chicken meats collected from poultry processing plants in nationwide. Korean J Microbiol 43:186-192.
- Zhu J, Wang Y, Song X, Cui S, Xu H, Yang B, Huang J, Liu G, Chen Q, Zhou G, Chen Q, Li F. 2014. Prevalence and quantification of *Salmonella* contamination in raw chicken carcasses at the retail in China. Food Control 44:198-202.
- Zwietering MH, van Gerwen SJC. 2000. Sensitivity analysis in quantitative microbial risk assessment. Int J Food Microbiol 58:213-221.